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# Nutritional Potential of Some Invasive Species of Macaronesia for Ruminants

C.S.A.M. Maduro Dias, C.F.M. Vouzela, H.J.D. Rosa, J.S. Madruga and A.E.S. Borba

University of the Azores, Faculty of Agricultural and Environmental Sciences, Institute of Agricultural and Environmental Research and Technology (IITAA). Rua Capitão João d'Ávila, 9700-042 Angra do Heroísmo, Açores, Portugal.

### ABSTRACT

Macaronesia islands' invasive plant use in animal feed or composting may bring economic and environmental benefits to the region. *Arundo donax*, *Pennisetum setaceum*, *Agave americana*, and *Ricinus communis*, present in the three archipelagos (Canary, Azores and Madeira), were characterized chemically and biologically. *A. donax* and *P. setaceum* showed elevated crude protein (CP) content, 13.25 and 16.33 DM%, respectively, and extremely high NDF values, 75.87 and 80.83 DM%, with a DM digestibility of 55.02 to 59.77%. *A. americana* showed a low NDF value (22.78 to 27.94 DM%) and a very low CP value (4.24 to 5.61 DM%). However, its DM digestibility was high (79.89 to 86.33%). *R. communis* presented the best values for CP (24.62%) and NDF (26.56 DM%), however, due to the presence of toxic substances (ricin), it cannot be easily used in animal feed. The *P. setaceum* and *R. communis* were found to be the least gas-producing forage, with *A. americana* being the major producer. To increase these plants' value for animal feed, treatment with urea or NaOH to *A. donax* and *P. setaceum*, and enrichment with nitrogen to *Agave* is proposed. Due to its toxic properties, *R. communis* must be used in composting.

**Keywords:** Invasive plants; Nutritive valorisation; Composting; Macaronesia

### \*Correspondence to Author:

A.E.S. Borba

University of the Azores, Faculty of Agricultural and Environmental Sciences, Institute of Agricultural and Environmental Research and Technology (IITAA). Rua Capitão João d'Ávila, 9700-042 Angra do Heroísmo, Açores, Portugal.

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## Introduction

Invasive plants are a problem that, to a greater or lesser extent, affect the region of Macaronesia, constituting a threat to the region's endemic flora and fauna. The use of these plant species as fibre sources for industry, animal feeding or composting may be considered as complementary means of combating their propagation, as suggested by Silliman *et al.* (2014)<sup>1</sup>.

The use of invasive plant species in animal feed has a long tradition in the Azores Islands. There are two clear periods of low grassland productivity, those being, summer (particularly August and September) and winter (from November through February). During winter, animals are usually kept in the so-called "invernadores", as an effort to protect them from adverse conditions, particularly, rain and wind (Borba *et al.*, 2015)<sup>2</sup>

During these periods of lack of grass, often unconventional fodders are used as sources of fibre, with shrubs playing a predominant role. Of the unconventional fodders used in Azorean animal feeding, we highlight *Pittosporum undulatum* (incense), *Hedychium gardnerianum* (ginger lily) (Borba *et al.*, 2015)<sup>2</sup>.

A review of literature indicates that there is limited information on the nutritive value of invasive plants. Some authors have carried out studies on the potential of invasive plants, namely on their chemical composition, nutritional value and toxicity (Zangerl and Berenbaum, 2005, Smith *et al.*, 2013, Burritt and Hart, 2014, Drossart *et al.*, 2017, Obour *et al.*, 2017)<sup>3,4,5,6,7</sup>. The findings of this analysis suggest that invasive plants are far more likely to cause significant impacts on resident plant and animal richness on islands than on the continent (Pyšek *et al.*, 2012)<sup>8</sup>. Some authors have reported that invasive plant contribution to ecosystem services is controversial due to the, mostly negative, relationship that these species have with the native flora. However, their continued dominance in many regions warrants a more thorough evaluation of their impact, both positive and negative, on the ecosystem (Gordon, 1998, Hershner and Havens, 2008)<sup>9,10</sup>.

The negative impact of non-native species to the loss of biological diversity (genetic, species, and ecosystem diversity) and the threat they represent to human health and welfare when they become invasive has been widely discussed in literature (Manchester and Bullock, 2000)<sup>11</sup>. Nonetheless, in certain instances, they can also provide conservation benefits. Furthermore, a fraction of non-native species will continue to cause biological and economic damage, as well as substantial uncertainty surrounding the prospective effects of all non-native species (Schlaepfer *et al.*, 2010)<sup>12</sup>. For example, their influence on the bee population remains quite unclear and is still a controversial matter of debate among researchers. Non-native species can currently contribute to the ecosystem's conservation objectives, which might include, for example, providing habitat or food resources to rare species, serving as functional substitutes for extinct taxa, or providing desirable ecosystem functions (Jordaan and Downs, 2012)<sup>13</sup>.

Invasion by exotic plants tends to be associated with the nutrient enrichment of soils. This phenomenon happens particularly on soils of naturally low fertility (Tabassum and Leishman, 2016)<sup>14</sup>, in which, invasive species may contribute to a homogenisation of soil conditions in invaded landscapes (Dassonville *et al.*, 2008)<sup>15</sup>.

Most of the invasive plants or weeds compete with desirable vegetation and adversely affect forage production and quality. These invasive species are often extremely destructive and difficult to control, allowing infestations to persist for several years and spread to new areas. Some plants have sharp spines, while others may lead to animal fatalities, either through direct poisoning or through an accumulation of nitrates and soluble oxalates (Scott and Robbin, 2005, Panter *et al.*, 2011)<sup>16,17</sup>.

The purpose of this study is the valorisation of natural fibres coming from vegetable invasive species in Macaronesia, such as *A. donax* L. (common cane), *Pennisetum setaceum* (known as fountain grass), *Agave americana* (similar to sisal) and *Ricinus Communis* (castor bean

plant), all of them propagated without control in the three Archipelagos (Canarias, Azores and Madeira). These plants are included in the International Union for Conservation of Nature (IUCN)'s TOP100 of most dangerous invasive species.

## Material and Methods

### Forage collection and preparation

The current study was conducted at the Animal Nutrition Laboratory, Department of Agricultural Sciences, University of the Azores, Azores, Portugal. Samples of plants were collected on Terceira and Santa Maria islands. This region is dominated by soils from basaltic lava mantle, known as "litolitic soils" according to Ricardo *et al.* (1979)<sup>18</sup>. They would fit in the Typic Udorthents according to Soil Taxonomy (USDA, 2014)<sup>19,20</sup>.

Samples were harvested manually, consisting of the parts of the plant animals normally eat.

### Chemical analysis

Dried samples were then ground through a 1-mm screen. These ground samples were analysed for dry matter (DM, method 930.15), crude protein (CP, method 954.01) and total ash method (942.05), according to the standard methods of AOAC (1995)<sup>21</sup>. Crude protein was determined by the standard micro-Kjeldahl method. Neutral detergent fibre (NDF), acid detergent fibre (ADF) and acid detergent lignin (ADL) were determined according to Goering and Van Soest (1970)<sup>22</sup>. *In vitro* digestibility was determined using the Tilley and Terry (1963)<sup>23</sup> method, modified by Alexander and McGowan (1966)<sup>24</sup>, and the juice of the rumen was obtained from a slaughterhouse, as described by Borba *et al.* (2001)<sup>25</sup>.

### In vitro Gas Production

*In vitro* gas production (GP) technique simulates the rumen fermentation process and it has been used to evaluate the potential of feeds to produce greenhouse gas. It is similar to the ruminal process, as gas (CO<sub>2</sub> and CH<sub>4</sub>) is produced from the carbohydrate fermentation.

Each assay was repeated three times (runs). Blanks were used for each inoculum to measure

the fraction of total gas production due to the substrate in inocula and these values were subtracted from the total to obtain the net GP. All treatments, for each assay, were incubated simultaneously in all runs, as per Menke *et al.* (1979)<sup>26</sup>.

Rumen digesta was collected as described by Borba *et al.* (2001)<sup>25</sup>. The preparation of buffer solutions and rumen inocula was as described by Menke and Steingass (1988)<sup>27</sup>.

The initial gas volume was recorded after 4, 8, 12, 24, 48, 72 and 96 hours of incubation.

This gas production represents the kinetic of the rumen's apparent GP and is expressed by the McDonald (1981)<sup>28</sup> equation. Gas production profiles were obtained after fitting the data to the exponential equation of Ørskov and McDonald (1979)<sup>29</sup>:

$$p = a + b (1 - \exp^{-ct})$$

Where:  $p$  represents the gas production at time  $t$ , the values of  $a$ ,  $b$  and  $c$  represent *constant* values in the exponential equation,  $a+b$  the total potential gas production (ml/g DM), and  $c$  the rate constant.

## Results and Discussion

From the results presented in Table 1, it was observed that *A. donax* and *P. setaceum* show elevated crude protein (CP) values, 13.25 and 16.33 DM%, respectively, and extremely high NDF values (75.87 and 80.83 DM%, respectively), which leads to a DM digestibility of 55.02 and 59.77%. The *A. americana*, although with a low NDF value (22.78 and 27.94 DM% for Terceira Island and Santa Maria, respectively), presents a very low CP value (4.24 and 5.61% for Terceira and Santa Maria, respectively), lower than the 7%, which is usually considered the minimum required value for the normal functioning of microorganisms (Lazzarini *et al.*, 2009)<sup>30</sup>. However, its dry matter digestibility is high (86.33 and 79.89% for Terceira and Santa Maria, respectively). The *R. communis* is, of all the studied samples, the one that shows the best values, 24.62 DM% for CP and 26.56 DM% for NDF.

**Table 1. Composition of the different sources of fibre.**

Treatment	DM (%)	100 g DM						DMD (%)	OMD (%)
		CP	NDF	ADF	ADL	EE	Ash		
<i>Arundo donax</i>	19.52	13.25	75.87	39.72	4.21	1.58	10.53	55.02	50.03
<i>Pennisetum setaceum</i>	18.70	16.33	80.83	41.90	4.77	1.28	16.30	59.77	51.99
<i>Agave americana</i> Terceira	14.88	4.24	22.78	20.04	5.50	1.69	4.49	86.33	85.43
<i>Agave americana</i> Santa Maria	10.09	5.61	27.94	24.42	4.39	1.45	12.06	79.89	77.15
<i>Ricinus communis</i>	19.27	24.62	26.56	20.02	4.38	2.30	9.59	78.06	76.34

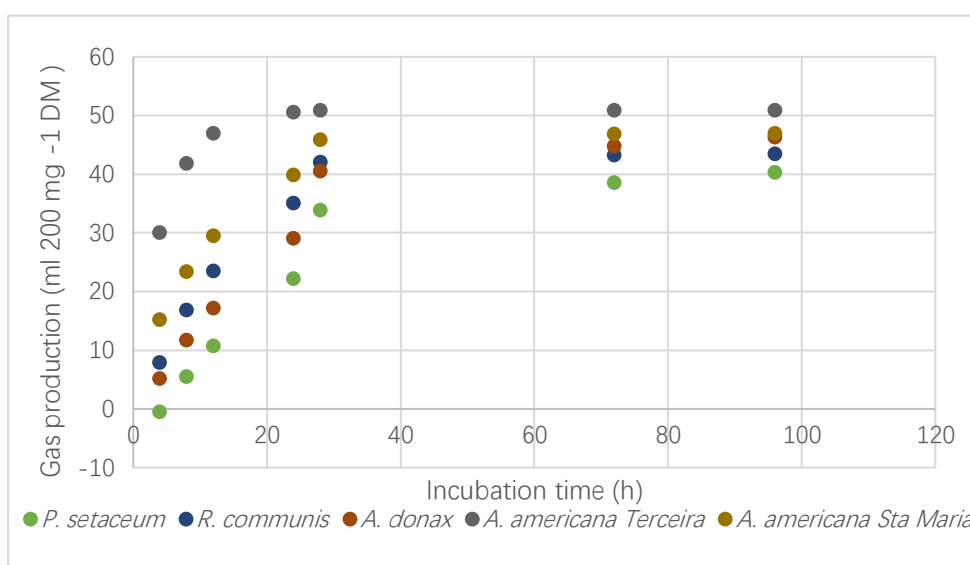
DM – Dry Matter, CP – Crude Protein, NDF – Neutral Detergent Fibre, ADF – Acid Detergent Fibre, ADL – Acid Detergent Lignin, EE – Extract Ether, DND –Dry Matter Digestibility, OMD –Organic Matter Digestibility.

Comparing the chemical composition values of *A. donax* found in this study with those reported by other authors, we found high fibre values (NDF) greater than 65% of DM and crude protein between 9.9% of DM and 11.1% of DM (Ahmed *et al.*, 2009 and Ahmed *et al.*, 2011, N.AG.RE.F., 2013)<sup>31,32,33</sup>. Regarding the digestibility of DM, other authors found values comparable to those

found in this study, between 47 and 52% (TagelDin, 1990, Ahmed *et al.*, 2011)<sup>33,34</sup>. Talapatra (1950)<sup>35</sup> reports a digestibility value of DM of 69% for *A. Donax* of India and N.AG.RE.F. (2013)<sup>32</sup> refers to a value of 66.8%. Baig and Bhagwat (2009)<sup>36</sup> and Behera *et al.* (2013)<sup>37</sup> report the galactopoietic properties of *A. donax* in dairy cows.

**Table 2. Equation terms for gas production, including residual standard deviations (rsd)**

	a	b	c	Lag Time (hr)	RSD
<i>Arundo donax</i>	-2.39	49.65	0.0419	1.2	1.24
<i>Pennisetum setaceum</i>	-7.56	49.1	0.0389	4.3	2.35
<i>Agave americana</i> Terceira	3.1	47.87	0.276	0	1.81
<i>Agave americana</i> Santa Maria	4.16	42.89	0.0749	0	0.99
<i>Ricinus communis</i>	-3.91	47.45	0.0722	1.2	1.38



**Figure 3. Pattern of *in vitro* gas production (fitted with exponential model) on incubation of invasive plants in buffered rumen fluid.**

Several authors argue that the palatability of *A. donax* is low, which results in a low voluntary intake, even when the animals ingest young plants (Ahmed *et al.*, 2011, Shehata *et al.*, 2006)<sup>33,38</sup>. The USDA (2014a)<sup>19</sup> reports that young *A. donax* plants are grazed, being one of the means of control of this weed during the dry season.

*P. setaceum* has an average nutritive value, an extremely high NDF value and a high crude protein value, however, unlike other *Pennisetum*, it is not normally used as a feed for ruminants. According to Joubert and Cunningham (2002)<sup>39</sup>, *P. setaceum* is an unpalatable species, possibly due to its serrated and rough leaves.

Fuentes-Rodriguez (1997)<sup>40</sup> research findings suggested that *Agave* generally has a low nutritional value in ruminant feed. With a low DM content (10%) and a low crude protein content (5% of DM), they are used both as an emergency maintenance feed and as part of the regular rations. Other authors reported values of 7.45% of crude protein in DM (Fraps, 1932 and Anon, 1942)<sup>41,42</sup>. These plants have good palatability and high humidity content (Suñigiga, 1980)<sup>43</sup>. For example, in Santa Maria, *Agave* is given to animals as a water source, in periods of forage shortage.

The literature presents chemical composition values for *Ricinus communis*, crude protein 23.7% between 22.5 and 24.8% of DM, NDF of 24.0%, ADF of 22.1% of DM and ADL of 2.8% of DM (Behl *et al.*, 1986, Bose *et al.*, 1988, Okorie *et al.*, 1985, Oorie and Anugwa, 1987, Purushotham *et al.*, 1985, 1986, Rao *et al.*, 1984)<sup>44,45,46,47,48, 49,50</sup>.

Although *Ricinus communis* is referred to as a toxic plant (Albuquerque *et al.*, 2014, Tokarnia *et al.*, 1975)<sup>51,52</sup>, some authors refer to the use of this plant in sheep feed. Lara *et al.* (2016)<sup>53</sup> reported values of chemical composition of *Ricinus communis* very similar to those found in this study (Table 1), that is, high values of crude protein (20.4% of DM), low NDF values (33.8% of DM) and a value digestibility of 76.8%. Barrales Heredia *et al.* (2018)<sup>54</sup> refer to the use of castor

oil in ruminant feed since it has a high crude protein content (62.6 to 66.77% of DM).

Invasive plant *in vitro* gas production results (Table 2) shows that the initial time of fermentation (Lag Time) varies greatly from forage to plant, ranging from 0 hours to 4.3 hours. This variation is in line with previous findings (Tuah *et al.*, 1996)<sup>55</sup>. It was observed that the *Agave* from Terceira and Santa Maria have a Lag Time of 0 hours, while the *Pennisetum setaceum* presents a Lag Time of 4.3 hours. According to the gas production curves (Figure 3), *Pennisetum setaceum* and *Arundo donax* are the least gas-producing plants.

According to the gas production curves (Figure 3), *P. setaceum* and *R. communis* are the least gas-producing plants, with *A. americana* from Terceira and Santa Maria being the major producers. We highlight the low potential of gas production of this fodder, and there were no results from other authors to compare those obtained in this work. However, in studies published by Moselhy *et al.* (2014)<sup>56</sup>, an inhibitory effect of two other invasive plants, *Pittosporum undulatum* and *Hedichium gardnerianum*, was verified in gas production.

## Conclusions

It is concluded that the studied invasive plants can be used as ruminant feed in a period of shortage of forages, or as a way to control their spread. The *Ricinus communis*, due to the presence of toxic substances in this plant, is not easily used in animal feed. As a strategy to increase the value of these plants, we propose that treatment with urea or NaOH is applied to *A. donax* and *P. setaceum*, and enrichment with nitrogen to *Agave*, for use in animal feed. Due to its toxic properties, *R. communis* must be used in composting.

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## References

1. Silliman, B.R., Mozdzer, T., Angelini, C., Brundage, J.E., Esselink, P., Bakker, J.P., Gedan, K.B., van de Koppel, J. and Baldwin, A.H. 2014. Livestock as a potential biological control agent for an invasive wetland plant. *Peer Journal*, 2: e567. doi: 10.7717/peerj.567
2. Borba, J.P.R., Maduro Dias, C.S.A.M., Rosa, H.J.D., Vouzela, C.F.M., Rego, O.A., Borba, A.E.S. 2015. Nutritional valorization of ginger lily forage (*Hedychium gardnerianum*, Sheppard ex Ker-Gawl) for animal feeding: treatment with urea. *African Journal of Agricultural Research*, 10:4606-4615. <https://doi.org/10.5897/AJAR2015.10075>
3. Zangerl, A.R. and Berenbaum, M.R. 2005. Increase in toxicity of an invasive weed after reassociation with its co-evolved herbivore. *Proceedings National Academy Sciences U S A*, 102:15529-15532. <https://doi.org/10.1073/pnas.0507805102>
4. Smith, S.B., DeSando, S.A. and Pagano, T. 2013. The Value of Native and Invasive Fruit-Bearing Shrubs for Migrating Songbirds. *Northeastern Naturalist*, 20:171-184. <https://doi.org/10.1656/045.020.0114>
5. Burritt, B., Hart, R.A. 2014. Nutritional Value and toxins in various noxious weeds. *All Current Publications. Paper 795*. [https://digitalcommons.usu.edu/extension\\_curall/795](https://digitalcommons.usu.edu/extension_curall/795).
6. Drossart, M., Michez, D. and Vanderplanck, M. 2017. Invasive plants as potential food resource for native pollinators: A case study with two invasive species and a generalist bumble bee. *Scientific Report*, 7: 16242. <https://doi.org/10.1038/s41598-017-16054-5>
7. Obour, R., Oppong, S. K. and Abebrese, I. K. 2017. Chemical Composition and Nutritive Value of an Invasive Exotic Species *Broussonetia Papyrifera* in Ghana. *Journal of Natural Sciences Research*, 7:45-53.
8. Pyšek, P., Jarošík, V., Hulme, P.E., Pergl, J., Hejda, M., Schaffner, U. and Vilà, M. 2012. *Glob Chang Biology*, 18: 1725–1737. doi: 10.1111/j.1365-2486.2011.02636.x
9. Gordon, D.R. 1998. Effects of invasive, non-indigenous pant species on ecosystem processes: Lessons from Florida. *Ecological Applications*, 8: 975–989.
10. Hershner, C. and Havens, K.J. 2008. Managing invasive aquatic plants in a changing system: strategic consideration of ecosystem services. *Conservation Biology*, 22:544-550.
11. Manchester, S.J. and Bullock, J.M. (2000). The impacts of non-native species on UK biodiversity and the effectiveness of control. *Journal of Applied Ecology*, 37: 845-864. <https://doi.org/10.1046/j.13652664.2000.00538.x>
12. Schlaepfer, R. M.A., Sax, D.F. and Olden, D. 2010. The Potential Conservation Value of Non-Native Species. *Conservation Biology*, 25: 428–437. DOI: 10.1111/j.1523-1739.2010.01646.x
13. Jordaan, L.A. and Downs, C.T. 2012. Nutritional and Morphological Traits of Invasive and Exotic Fleshy-fruits in South Africa. *Biotropica*, 44:738-743. *Conservation Biology*, 25: 428–437. DOI: 10.1111/j.1523-1739.2010.01646.x
14. Tabassum, S. and Leishman, M.R. 2016. Trait values and not invasive status determine competitive outcomes between native and invasive species under varying soil nutrient availability. *Austral Ecology*, 41: 875-885. <https://doi.org/10.1111/aec.12379>
15. Dassonville, N., Vanderhoeven, S., Vanparys, V., Hayez, M., Gruber, W. and Meerts, P. 2008. Impacts of alien invasive plants on soil nutrients are correlated with initial site conditions in NW Europe. *Published in: Oecologia*, 157: 131-140
16. Scott, L. and Robbins, K. 2005. Invasive plants that are toxic to livestock. *Ministry of Agriculture and Lands. British Columbia, Canada.*
17. Panter, K.E., Ralphs, M.H., Pfister, J.A., Gardner, D.R., Stegelmeier, B.L., Lee, S.T., Welch, K.D., Green, B.T., Davis, T.Z. and Cook, K. 2011. *Plants Poisonous to Livestock in the Western States. U.S. Department of Agriculture, Agricultural Research Service, Poisonous Plant Research Laboratory, Logan, Utah.*
18. Ricardo, R. P., Madeira, M. A. V. and Medina, J. M. B. 1979. Enquadramento taxonómico dos principais tipos de solos que se admite ocorrerem no Arquipélago dos Açores. *Anais do Instituto Superior de Agronomia*, 38: 167-180.
19. USDA 2014a. Field guide for managing Giant reed in the Southwest. *USDA Forest Service, Albuquerque, USA.*
20. USDA -United States Department of Agriculture 2014. *Keys to Soil Taxonomy. Twelfth Edition.*
21. AOAC - Association of Official Analytical Chemists 1995. *Official Methods of Analysis. 16th ed. Association OF Official Analytical Chemists, Virginia, USA.*
22. Goering, H.K. and Van Soest, P.J. 1970. Forage fiber analyses. *Agricultural Handbook nº379, Washington, DC, USA.*
23. Tilley J.M.A. and Terry, R. A. 1963. A two-stage technique for the in vitro digestion of forage crops. *Journal of the British Grassland Society*, 18: 104-111. doi/abs/10.1111/j.1365-2494.1963.tb00335.x
24. Alexander RH, McGowan M 1966. The routine determination of in vitro digestibility of organic matter

- in forages. An investigation of the problems associated with continuous large-scale operation. *Journal of the British Grassland Society*, 21: 140 – 147. doi/10.1111/j.1365-2494.1966.tb00462.x
25. Borba, A.E.S., Correia, P.J.A., Fernandes, J.M.M., Borba, A.F.R.S. 2001. Comparison of three sources of inocula for predicting apparent digestibility of ruminant feedstuffs. *Animal Research*, 50:265-274. <https://doi.org/10.1051/animres:2001113>
  26. Menke, K.H., Raab, L., Salewski, A., Steingass, H., Fritz, D. and Schneider, W. 1979. The estimation of the digestibility and metabolizable energy content of ruminant feedstuffs from the gas production when they are incubated with rumen liquor in vitro. *Journal of Agricultural Science, Cambridge*, 92: 217-222. <https://doi.org/10.1017/S0021859600086305>
  27. Menke, K.H. and Steingass, H. 1988. Estimation of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. *Animal Research and Development* 28: 7–55.
  28. McDonald I 1981. A revised model for the estimation of protein degradability in the rumen. *Journal of Agricultural Science, Cambridge*, 96: 251-252. <https://doi.org/10.1017/S0021859600032081>
  29. Ørskov, E.R. and McDonald, P. 1979. The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *Journal of Agricultural Science, Cambridge*, 92:499-503.
  30. Lazzarini, I., Detmann, E., Sampaio, C.B., Paulino, M.F., Valadares Filho, S.C., Souza, M.A. and Oliveira, F.A. 2009. Dinâmicas de trânsito e degradação da fibra em detergente neutro em bovinos alimentados com forragem tropical de baixa qualidade e compostos nitrogenados. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 61: 635-647. <http://dx.doi.org/10.1590/S010209352009003000017>
  31. Ahmed, M. E., Shehata, E. I., Ammou, F. F. A., Khalifa, E. I. and El-Zolaky, O. A. 2009. Productive and reproductive performance of Rahmani sheep fed rations containing reed forage (*Arundo donax* L.) either fresh, hay or silage. *Egyptian J. Sheep Goat Sci.*, 4: 45-54.
  32. N.AG.RE.F. – National Agricultural Research Foundation 2013. Fibre Crops as source for Animal Feeding. Fibre Summers School. Euphratis School, Catania, Greece. 25/7/2013.
  33. Ahmed, M. E., El-Zelaky, O. A., Aiad, K. M. and Shehata, E. I. 2011. Response of small ruminants to diets containing reed forage either as fresh, silage or hay versus berseem hay. *Egyptian J. Sheep & Goat Sci.*, 6: 15-26.
  34. Tagel-Din, A. E. 1990. Evaluation of reeds in complete diets for ruminant animals. *Indian Journal Animal Science*, 60: 1106-1109
  35. Talapatra, S. K. 1950. The nutritive value of the indigenous grasses of Assam. III. The semi-aquatic grasses as cattle feeds. *Indian Journal Veterinary Science*, 20: 229-240
  36. Baig, M. I. and Bhagwat, V. G. 2009. Study the efficacy of GALACTIN VET bolus on milk yield in dairy cows. *Vet. World*, 2: 140-142
  37. Behera, P. C., Tripathy, D. P., Parija, S. C. 2013. Shatavari: potentials for galactagogue in dairy cows. *Indian J. Trad. Knowledge*, 12: 9-17.
  38. Shehata, E. I., Ahmed, M. E., Ammou, F. F. A., Soliman, A. A. M., Aiad, K. M. and Abdel-Gawad, A. M. 2006. Comparison of feeding reed as hay or silage with feeding berseem hay or maize silage to dairy Zaraibi goat. *Egyptian Journal Sheep, Goat and Desert Animal Science*, 1: 233
  39. Joubert, D.F. and Cunningham, P.L. 2002. The distribution and invasive potential of Fountain Grass *Pennisetum setaceum* in Namibia. *Dinteria*, 27: 37-47.
  40. Fuentes-Rodriguez J. 1997. A comparison of the nutritional value of Opuntia and Agave Plants for ruminants. *Journal of the Professional Association for Cactus Development*, 2:20-24.
  41. Fraps, G. S. 1932. The composition and utilization of Texas feeding stuffs. Texas Agricultural Experiment Station. Bulletin No. 461
  42. Anon 1942. Analyses of Rhodesian foodstuffs. *Rhodesia Agric. J.*, 39: 391-398.
  43. Suñigiga, C. H. 1980. Utilización del Maguey como Suplemento en el Crecimiento de Becerras Holstein. ITESM. Monterrey, NL. Mexico.
  44. Behl, C. R., Pande, M. B., Pande, D. P., Radadia, N. S. 1986. Nutritive value of matured wilted castor (*Ricinus communis* Linn.) leaves for crossbred sheep. *Indian J. Anim. Sci.*, 56: 473-474
  45. Bose, M. L. V., Wanderley, R. da C., Carvalheira Wanderley, R. 1988. Digestibility of detoxified castor meal and lucerne hay in sheep and metabolism of nitrogen fraction. *Rev. Soc. Bras. Zootec.*, 17: 456-464.
  46. Okorie, A. U. and Anugwa, F. O. I. 1987. The feeding value of roasted castor oil bean (*Ricinus communis*) to growing chicks. *Plant Foods Human Nutrition*, 37: 97-102. doi:10.1017/S0021859600063048
  47. Okorie, A. U., Anugwa, F. O. I., Anamelechi, G. C. and Nwaiwu, J. 1985. Heat treated castor oil bean (*Ricinus communis*): a potential livestock protein supplement in the tropics. *Nutrition Reproduction International*, 32: 659-666.



48. Purushotham, N. P., Veeraraghavan, G., Naidu, M. M. and Mahender, M, 1985. Haematological studies on experimental feeding of castor bean meal (*Ricinus communis*) in sheep. *Indian Veterinary Journal*, 62: 379-382.
49. Purushotham, N. P., Rao, M. S. and Raghavan, G. V. 1986. Utilization of castor beanmeal in the concentrate mixture of sheep. *Indian Journal of Animal Science*, 56: 1090-1093.
50. Rao, M. S., Purushotham, N. P., Raghavan, G. V. and Mahender, M. 1984. Biochemical changes in experimental feeding of castor bean meal (*Ricinus communis*) in sheep. *Indian Journal Veterinary Pathology*, 8: 33-36.
51. Albuquerque, S. S. C., Rocha, B. P., Albuquerque, R. F., Oliveira, J. S., Medeiros, R. M. T., Riet-Correa, F., Evêncio-Neto, J., Mendonça, F. S. 2014. Intoxicação espontânea por *Ricinus communis* (Euphorbiaceae) em bovinos. *Pesquisa Veterinária Brasileira*, 34:827-831.
52. Tokarnia, C.H., Dobereiner, J., Canella, C. 1975. Intoxicação experimental em bovinos pelas folhas de *Ricinus communis*. *Pesquisa Agropecuária Brasileira*, 10: 1-7.
53. Lara, C., Del Viento, A. and Palma, J.M. 2016. Preferencia y consumo de diferentes partes morfológicas del *Ricinus communis* L. (higuerilla) por ovinos. *Avances en Investigacion Agropecuaria*, 20: 43-52.
54. Barrales Heredia, S.M., Ochoa Landin, M.E., Barrera Silva, M. A., Anaya Islas, J., Huez López, M.A. 2018. Utilización de pasta de higuerilla (*Ricinus communis* L.) como suplemento alimenticio para ganado. In: XXVI Producción de carne y leche en climas cálidos: Reunión Internacional. UABC Mexicali - Baja California – México.
55. Tuah, A K, Okai, D. B., Ørskov, E.R., Kyle, D., Wshand, Greenhalgh, J. F. D., Obese, F. Y. and Karikari, P. K. 1996. In sacco dry matter degradability and in vitro gas production characteristics of some Ghanaian feeds. *Livestock Re-search Rural Development*, 8 article 3, Retrieved June 26, 2018, from <http://www.lrrd.org/lrrd8/1/tuah.htm>
56. Moselhy, M.A., Nunes, H.P. and Borba, A.E.B. 2014. Effect of replacement of ordinary ruminant feed with *Hedychium gardnerianum* or *Pitosporum undulatum* on in vitro rumen fermentation characteristics. *International Journal of Advanced Research*, 2: 91-104.

