



## Journal of Pharmaceutical Research and Reviews (ISSN:2576-8417)



# Anti-cancer activity of quercetin via apoptosis induction pathways in human breast cancer cell lines-a systematic review and meta-analysis

Rawdha Benziane; Bunmi Ibrahim\*

School of Allied Health Sciences. De Montfort University, Leicester. United Kingdom

### ABSTRACT

A systematic review and meta-analysis were conducted to summarize and review the current literature surrounding quercetin in breast cancer and evaluate its efficacy as an anticancer agent in human breast cancer cell lines. Electronic databases, PUBMED, WEB OF SCIENCE and MEDLINE were systematically searched in December 2020 using the key terms, Quercetin AND Breast cancer AND apoptosis OR cell cycle. Fourteen papers surrounding effects of quercetin on cell viability were obtained. Risk of bias assessment revealed that most papers were reliable. Three meta-analyses were conducted to confirm the efficacy of quercetin at the concentrations of 40-50  $\mu$ M, 100 $\mu$ M, and >100 $\mu$ M (120- 200  $\mu$ M). Two subgroup analyses on incubation time and cell type were performed to ascertain whether these factors affect mechanisms of quercetin. Results showed that the decrease in percentage cell viability is directly proportional to increasing concentration. The greatest decrease in cell viability was observed in 100  $\mu$ M (Risk Ratio (RR) = 0.56;  $p < 0.00001$ ), followed by 100  $\mu$ M (RR= 0.74) and then 40-50  $\mu$ M (RR=0.79). However, effects of quercetin may be determined by other factors such as incubation time or a particular cell type signaling cascade. Subgroup analysis revealed that significant differences between 24 and 48 hours were not observed. MCF7 cells showed to be most sensitive to actions of quercetin ( $p < 0.0001$ ). It was concluded that there is enough information surrounding the cytotoxic effects of quercetin to be progressed forward, however optimal doses that are physiologically relevant and safe in humans should be elucidated.

**Keywords:** quercetin, breast cancer cells, apoptosis, meta-analysis, systematic review, cell viability

### \*Correspondence to Author:

Bunmi Ibrahim

School of Allied Health Sciences.  
De Montfort University, Leicester.  
United Kingdom

### How to cite this article:

Rawdha Benziane, Bunmi Ibrahim.  
Anti-cancer activity of quercetin via  
apoptosis induction pathways in  
human breast cancer cell lines-a  
systematic review and meta-analy-  
sis. Journal of Pharmaceutical Re-  
search and Reviews, 2022; 6:23.



eSciPub LLC, Houston, TX USA.

Website: <https://escipub.com/>

## Introduction

Breast cancer is the most prevalent cancer worldwide and is the leading cause of cancer related death in women [2], in which its most common treatment plans; chemotherapy and radiotherapy, exert side effects that are often unbearable to the patient and are highly susceptible to resistance [82][90].

The usage of natural products as anticancer agents has received great attention in recent times given the reported chemo preventive and chemotherapeutic qualities they possess [18]. Plant-derived anticancer drugs such as taxanes and vinca alkaloids already exist and are widely used in clinical settings which highlights the potential of plants as anti-cancer drugs [29]. These natural products would be highly beneficial as they are expected to be widely accessible, less processed, and less susceptible to resistance [18]. Flavonoids from plants are largely distributed across the common diet and have been reported to possess a broad range of health benefits including antimicrobial, anti-inflammatory, chemoprevention, and chemotherapeutic effects [6][41][42][61][70]. A major flavonoid, part of the flavanol subclass, quercetin has been shown to induce anticancer effects through, loss of cancer cell viability, apoptosis, and reduced cell proliferation which is determined via the role of quercetin in cytotoxic pathways [71].

A crucial hallmark of cancer, evasion of apoptosis is an attractive cancer therapy target, as it decelerates and halts the progression of cancer by re-sensitizing the cancer cells to undergo apoptosis [31][36][63]. Additionally, resistance to chemotherapy and radiotherapy has been attributed to this apoptotic defect, hence scrutinizing novel therapies in rectifying dysregulated apoptosis is important to solve the prominent aspect of resistance to treatment [36][63]. Quercetin has been widely reported to induce dose and time-dependent apoptosis and inhibit proliferation in various cancers including lung, colon, breast, prostate, and ovarian

cancers without affecting the tissue's normal counterparts [7][33][44][45].

Nonetheless, reports regarding quercetin and its roles in apoptotic pathways and cell cycle arrest have been inconsistent. Many studies report the induction of apoptosis by quercetin via the mitochondrial pathway [80] but others mention that quercetin upregulates the expression of members of the class O of forkhead box transcription factors (FOXO), which induces both mitochondrial dependent and independent apoptosis, in an MDA-MB-231 cell line [89][22]. Furthermore, other studies involving non-cancer cells such as damaged rat neurons demonstrated that low doses quercetin induced anti-apoptotic effects instead [28]. Additionally, there have been inconsistencies in the literature regarding the concentration at which quercetin is the most effective where some report statistical significance in concentrations as low as 10µM [21] whereas others demonstrate an effect seen only after 50µM [13]. Interestingly there has been some reports insinuating that quercetin promotes cancer cell growth at low concentrations [87].

Hence, these findings warrant a need to explore the activity of quercetin in breast cancer to determine whether there is a particular signaling cascade induced by quercetin and if this may be influenced by other factors such as dosage and cell type.

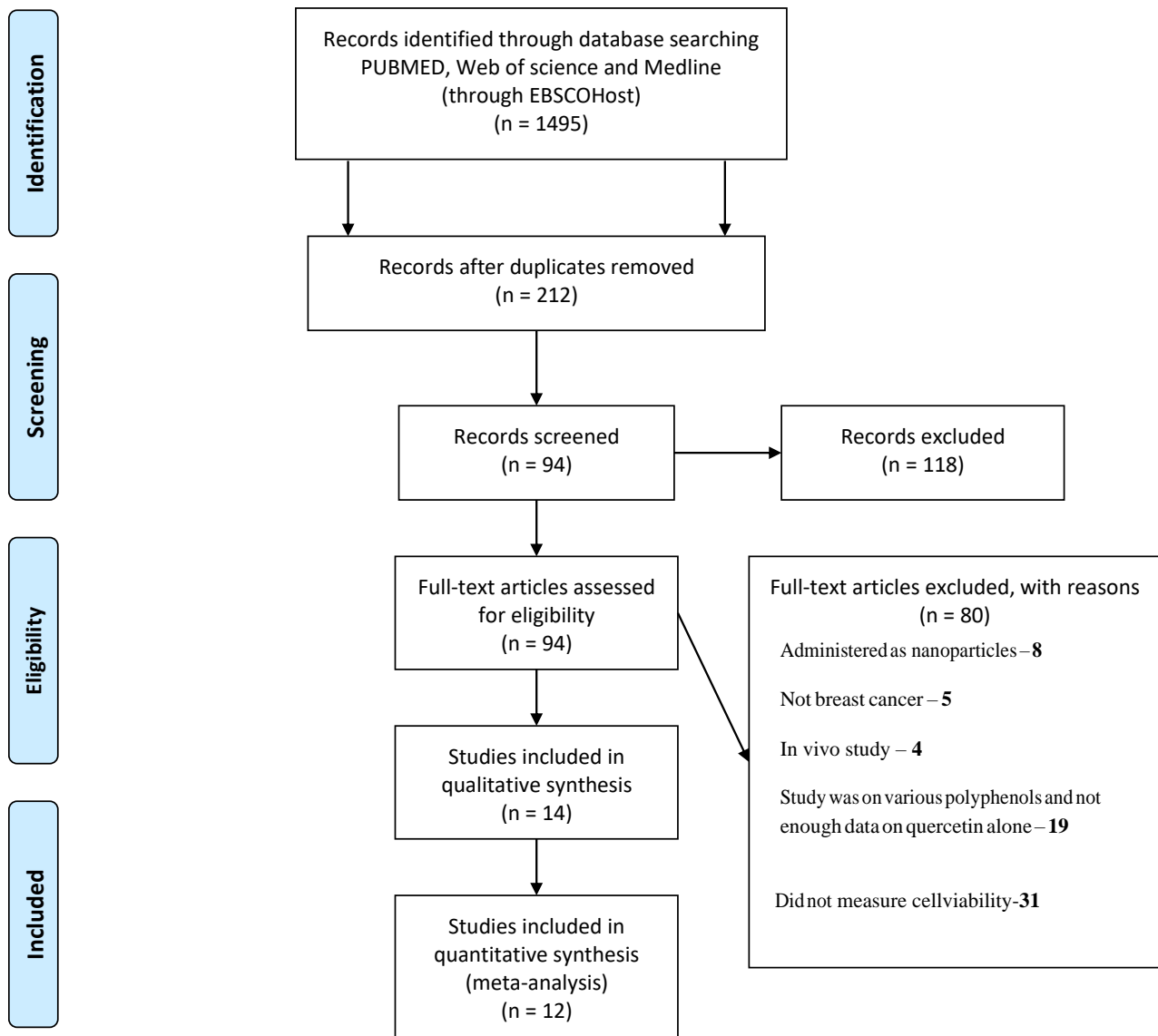
Moreover, despite the abundance of studies demonstrating the significant cytotoxic effects of quercetin in various human breast cancer cell lines the progression to human clinical trials is scarce and even animal *in vivo* studies are rather limited, to further consolidate these promising findings clinical application is required. This may be due to the lack of confidence in the pharmacokinetics of flavonoids, specifically quercetin where many reports its lack of bioavailability due to poor absorption and very rapid metabolism [11]. However, many have demonstrated that this challenge can be overcome via the manipulation of the delivery of quercetin, where successful administration is

seen through nanoparticles [11][72]. In spite of this, a phase 1 clinical trial has demonstrated that quercetin was able to achieve plasma levels which can inhibit tyrosine kinase activity [26]. This is further reinforced by the fact that more emerging studies have demonstrated a considerable level of diet- derived quercetin can be absorbed through the digestive tract [57].

It may be more economical, time-efficient, and safer to use quercetin conventionally by finding its optimal dosage in humans, as reports have shown increased risks of toxicities using novel drug delivery systems such as incomplete capsule degradation and organ accumulation

[11]. This makes it important to review the optimal dose in cellular studies and possibly inform future human clinical trials.

Consequently, a systematic review and meta-analysis was conducted to summarise and review the current literature surrounding quercetin in breast cancer and determine its efficacy as an anticancer agent in various human breast cancer cell lines. In all, this was done with the aim to conclude whether it is worth further investigation in clinical settings and if this flavonoid can be implemented into future breast cancer treatment regimes.



**Figure 1:** Flow chart of study selection process

## Materials and methods

### Search Strategy:

The population, intervention, controls, and outcomes (PICO) search strategy was employed for this systematic review. Papers based on human breast cancer cell lines (P) treated with quercetin (I) compared to a control (C) reporting an outcome on cell viability upon quercetin treatment which in turn was used as an indicator of apoptosis (O) were obtained (Figure 1).

The electronic databases: PUBMED, WEB OF SCIENCE and MEDLINE were searched in December 2020, using advanced search Booleans. Search terms were inputted as, Quercetin AND Breast cancer AND apoptosis OR cell cycle. MESH terms in PUBMED were also used and inputted as; ("Apoptosis"[Mesh]) AND "Breast Neoplasms"[Mesh]) AND "Quercetin"[Mesh] AND (flavanol)). All collected papers were saved in a Microsoft Excel spreadsheet where duplicates were then removed.

### Data Extraction:

Data was extracted from each paper and relevant information was inputted into a table (table 1). Data to be inputted into RevMan 5 was extracted from each study using the software WebplotDigitizer, where screenshots of graphs and bar plots were taken and loaded into this software.

### Statistical Analysis:

Quantitative analysis was undertaken via the completion of meta-analyses using the statistical software RevMan 5. This meta-analysis aimed to measure the significance of the effects of quercetin treatment on cell viability. Percentage cell viability was treated as dichotomous data where a risk ratio and confidence intervals were calculated. A random effects model was employed as variations between studies were bound to occur, therefore this model calculated the mean of distribution of effects, hence giving a more accurate representation of the actual impact of quercetin. Heterogeneity was calculated through the  $I^2$  index. To determine sources of heterogeneity two subgroup analyses were conducted on variables suspected to affect the functional outcomes of quercetin on cell viability. The first being a subgroup analysis on incubation time, the second was cell type.

### Risk of Bias

The evaluation of study methodologies was employed through answering the eligibility criteria of the in vitro section in the ToxRtool and categorizing studies based on reliability (Table 1).

### Results

In all, 14 papers were evaluated, in which 12 were included in the quantitative analysis to evaluate the impact of quercetin on percentage cell viability. Although 12 papers were included, some did test more than one cell type, concentration and incubation time and were therefore considered as separate studies.

**Table 1:** Risk of bias assessment of each study using *In Vitro* section of ToxRTool

Study	1	2	3	4	5	6	7	8	9	10	11	12	13	14
	[93]	[13]	[47]	[94]	[64]	[78]	[87]	[49]	[16]	[20]	[21]	[33]	[43]	[58]
<b>Study Criteria</b>														
Is test Substance identified?	1	1	1	1	1	1	1	2	1	1	1	1	1	1
Is origin of test Substance described?	1	1	1	1	1	3	1	1	1	1	1	1	3	1
Is test system sufficiently	1	1	1	2	1	1	1	1	1	1	1	1	1	1

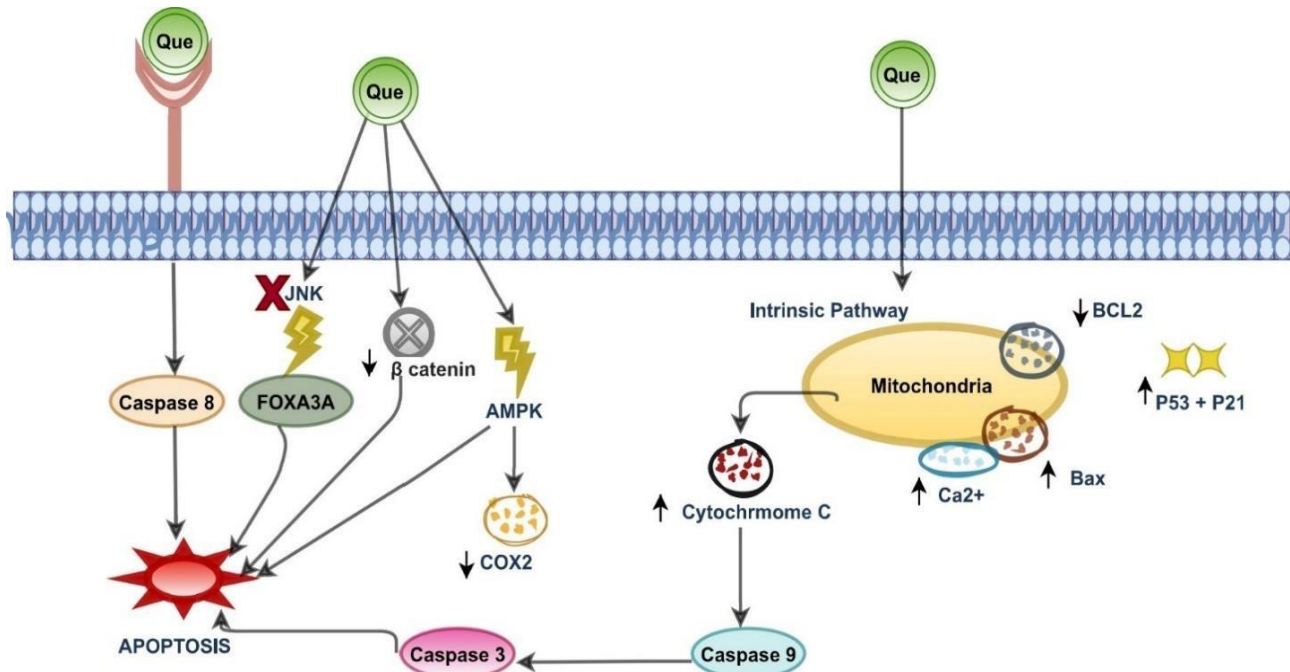
described?														
Is the method of test system given?	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Was a reference on the test system method given or is method generally known?	1	1	1	1	1	1	1	2	1	1	1	1	1	1
Is Necessary information regarding test system properties and conditions given?	1	1	1	1	1	1	1	1	1	1	1	1	2	1
Are concentrations administered given?	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Is duration and time of exposure described?	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Were variables which may impose secondary effects and influence results given ex: solubility/temperature?	2	2	2	2	2	2	2	2	2	2	2	2	2	2
Was a negative control included?	1	3	3	1	1	1	1	4	1	1	1	4	1	1
Was a positive control included?	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Is number of replicates given?	1	1	1	1	1	1	4	1	1	1	1	1	3	1
Is description of results complete and have all variables described in methods been described in results?	1	1	1	1	1	1	1	1	1	1	1	1	3	1
Has statistical analysis been described completely?	1	1	1	1	1	1	1	1	1	1	1	1	2	1
<b>Total Average</b>	1.07	1.07	1.21	1.21	1.07	1.21	1.07	1.21	1.21	1.07	1.07	1.28	1.64	1.07
<b>Category</b>	1	1	1	1	1	1	1	1	1	1	1	1	2	1

**Category 1** => Reliable without restriction, **Category 2** => Reliable with Restriction, **Category 3** => Not reliable, **Category 4** => Not assignable

<b>Table 2 - Summarised characteristics of each included study</b>							
<b>Author</b>	<b>Location</b>	<b>Cell Type</b>	<b>Detection Method</b>	<b>Cell Number</b>	<b>Treatment</b>	<b>Findings on cell Viability</b>	<b>Findings on cytotoxicity</b>
<b>Chien et al., 2009</b>	China	MDA-MB-231 cells	viable cells were determined by MTT	(2 x 10 <sup>5</sup> cells/well; 12- well plates)	0, 50, 100, 150, 200, 250 and 300 µM for 24h or 48h	Quercetin significantly decreased the percentage of viable cells – effects were dose dependent - (50% inhibition of cell growth, 24 h) was 278 µM	Apoptosis induced by the mitochondrial pathway
<b>Chou et al., 2010</b>	Taiwan	MCF-7 cells	Trypan blue exclusion and PI-exclusion	(2 x 10 <sup>5</sup> cells/well; 12- well plates)	0, 10, 50, 100, 150 and 175 µM quercetin and was incubated for 24 and 48 h	Quercetin decreased the percentage of viable cells - effects were dose and time dependent - (50% inhibition of cell growth, 48 h) was 92.4 µM	Apoptosis induced by the mitochondrial pathway
<b>Deng et al., 2013</b>	China	MCF 7 Cells	viable cells were determined by MTT	(5x10 <sup>3</sup> /well) were plated in 96-well plates	0, 2.5, 5, 10, 20 and 40 mg/ml after 24 or 48 h	highest inhibition rate was 58.72% and the rate of inhibition was concentration- and time- dependent.	The apoptosis rate of the quercetin 40 mg/ml group was 37.81%, which was higher than the apoptosis rates in the low concentration (20mg/ml) and control groups. surviving mRNA levels were reduced when the concentration of quercetin increased.
<b>Devipria et al., 2015</b>	India	MCF 7	viable cells were determined by MTT	(1 x 10 <sup>6</sup> ) were plated in 96- well plates	0, 10, 20, 30, 40, 50, 60 ug/ml quercetin and incubated for 24, 48 and 72 h	quercetin decreased the percentage of viable cells in dose and time dependent manner. Incubation for 72 h resulted in complete loss of cells	Dose and time dependent decrease in cytosolic calcium
<b>Dhumale et al., 2015</b>	India	MCF 7	Trypan Blue exclusion	(1 x 10 <sup>6</sup> ) were plated in 96-well plates	0,10,25, 50 µM quercetin incubated for 6, 12, 24, 48 h	Dose and time dependent cell viability decrease	MCF-7 cells treated with 50 µM of quercetin showed massive cell death at 6 h onward
<b>emzaei et al., 2017</b>	Iran	MCF7	Viable cells were determined by MTT	(1 x 10 <sup>6</sup> ) were plated in 96- well plates	10, 20, 40, 80 and 120 µM incubated for following 24, 48 and 72 h	Cancer cell growth inhibition was dose and time dependent (50% inhibition, 24 hours) was 105.4uM + (52.5uM for 48 h)	
<b>Jeong et al., 2010</b>	USA	SK-BR3	Trypan Blue Exclusion	Not stated	100 µM incubated for 48h		level of Her-2/neu protein began to decrease after 8 h of quercetin treatment + quercetin also dephosphorylated PI3K and Akt

<b>Khorsandi et al., 2017</b>	Iran	MCF7 cells	Viable cells were determined by MTT	number not stated - MCF-7 left culture media for 48 h in 24 well plates	50 $\mu$ M/ml for 48 h	Percentage of viable cells significantly decreased	The proliferation of MCF-7 cells was significantly decreased. Apoptosis induced by mitochondrial pathway
<b>Kiyga et al., 2020</b>	Turkey	MCF7 cells + MDAMB 231	Viable cells were determined by MTT	$1 \times 10^4$ and $1.5 \times 10^4$ cells/well	10, 25, 100 $\mu$ M for 48 h.	Increase in cell death in a dose and time dependent manner + MCF-7 cells were shown to be more sensitive than MDA-MB-231 cells	Apoptosis was induced in treated cells with 25 and 100 $\mu$ M. Higher level of caspase-activity was detected in MCF-7 cells - pro-caspase-3 was expressed in the MCF-7 cell lines
<b>Lee et al., 2009</b>	Korea	MCF7	Viable cells were determined by MTT	seeded on 96-well microplates at 4,000 cells/well	25, 50, 100, 200, or 400 $\mu$ M for 6 h	Decrease in cell viability was in a dose dependent manner	Apoptotic cell death was increased with 100 $\mu$ M quercetin treatment, as shown with chromatin condensation + Quercetin strongly activated AMPK and increased P53 + P51
<b>Nguyen et al., 2017</b>	Korea	MDAMB231	Viable cells were determined by MTT	$1 \times 10^4$ in 96 well-plates	2.5-80 $\mu$ M for 24 h, 48 h and 72 h	Decrease in cell viability was in a dose dependent manner	quercetin caused cell cycle arrest at S and G2/M phase + of Foxo3a activity + JNK inhibition
<b>Prandhan et al., 2015</b>	India	MCF7 cells	Viable cells were determined by MTT	$1 \times 10^4$ in 96 well-plates	25, 50, 75, or 100 $\mu$ M for 24h	quercetin inhibited cell viability after 24h of treatment	Apoptosis induced by mitochondrial pathway Cells became elongated, losing their characteristic morphology
<b>Sultan et al., 2017</b>	Egypt	MDA-MB-157 and MDA-MB-231	Viable cells were determined by MTT	stock cell suspension containing 20,000 cells/ml. 100 $\mu$ l were seeded per well of 96-well plate	0-550 $\mu$ M for 48 h	significant dose-dependent cytotoxic effects on cells when compared to control cells + MDAMB231 => more sensitive than MDAMB157	Quercetin induced apoptosis of both treated cell lines in a caspase- dependent manner + downregulation of FASN + B catenin
<b>Xu et al., 2020</b>	China	MCF-7 cells MDA-MB-231	CCK-8 assay and Realtime cell impedance analyser (RTCA) assay	96-well microplates ( $1 \times 10^4$ cells/well)	0.1-500 $\mu$ M with for 24-72 h	inhibited cell viability at higher concentrations ( $\geq 50 \mu$ M)	inhibited COX-2 protein expression+ p300 HAT-mediated acetylation of NF- $\kappa$ B p50, resulting in a marked reduction in acetyl-p50 protein levels



**Figure 2** Illustration demonstrating the reported apoptotic pathways induced by Quercetin

**Fig 2** - Illustration showing apoptotic pathways upon quercetin treatment. Apoptosis has been shown to be induced via the mitochondrial pathway where levels of pro-survival proteins BCL2 have been seen to decrease whereas pro apoptotic proteins increase when quercetin (Que) is administered [93][13][21][43][47][64][78] (fig 9). Others have demonstrated apoptosis through the extrinsic pathway [58]. Quercetin increases in Forkhead box O3 (FOX3a) proteins through c-Jun N-terminal kinase (JNK) pathway inhibition which have also been reported to induce apoptosis [58]. Another report suggests decrease in B catenin proteins inhibits in wnt signaling pathway and results in apoptosis [87]. The 5' AMP-activated protein kinase (AMPK) pathway has also been shown to be essential for apoptosis induction by quercetin [49].

### Meta-Analysis Results:

**Table 1:** Summarised Meta-Analysis Results Using Various Concentrations of Quercetin

Concentration	Risk Ratio (RR)	P value	I <sup>2</sup> index (heterogeneity)
<b>50 µM</b>	0.77 [0.71,0.85]	P<0.0001	91%
<b>100 µM</b>	0.74 [0.66,0.83]	P<0.0001	95%
<b>&gt;100 µM (120-200 µM)</b>	0.56 [ 0.44, 0.71]	P<0.0001	95%

The concentrations of 40-50 µM were chosen to initially determine the effects of quercetin as these were the lowest common doses across studies. Twelve studies were included in this analysis [13][21][47][47][33][58][49][64][78][87]. Cell viability in this section ranged from 46-93 %. Meta-analysis of the pooled results showed an RR

value of 0.77 [0.71- 0.85] with a P value of P< 0.00001; demonstrating a significant decrease in cell viability. Heterogeneity for this analysis had an I<sup>2</sup> value of 91% (additional data are given in Online resource 1). To determine most effective dosage quercetin at higher concentrations of 100µM (additional data are given in Online



resource 1) and  $>100\mu\text{M}$  were chosen. Results showed concentrations  $>100\mu\text{M}$  had the greatest effect on cell viability RR value being, 0.56,  $p<0.0001$  (additional data are given in Online resource 1).

#### Subgroup Analysis on Incubation Time:

Sub-group analyses were performed on potential factors which may have impacted the function of quercetin. A sub-group analysis was conducted on the various incubation times; 6, 24

and 48 hours (additional data are given in Online resource 1). Seven studies were in the 24- hour subgroup [93][13][47][64][87] and nine [103][13][94][47][78][87] in the 48- hour group; only one study was in the 6- hour subgroup [49]. Results obtained from the meta-analysis of the subgroups demonstrated that 48 hours resulted in an RR value of 0.67 [0.56,0.79];  $p<0.00001$  and this had the greatest effects on percentage cell viability compared to 24 hour and 6-hour incubation (additional data are given in Online resource 1).

**Table 2:** Summarized Subgroup Meta -analysis Results – Incubation Time = Subgroup

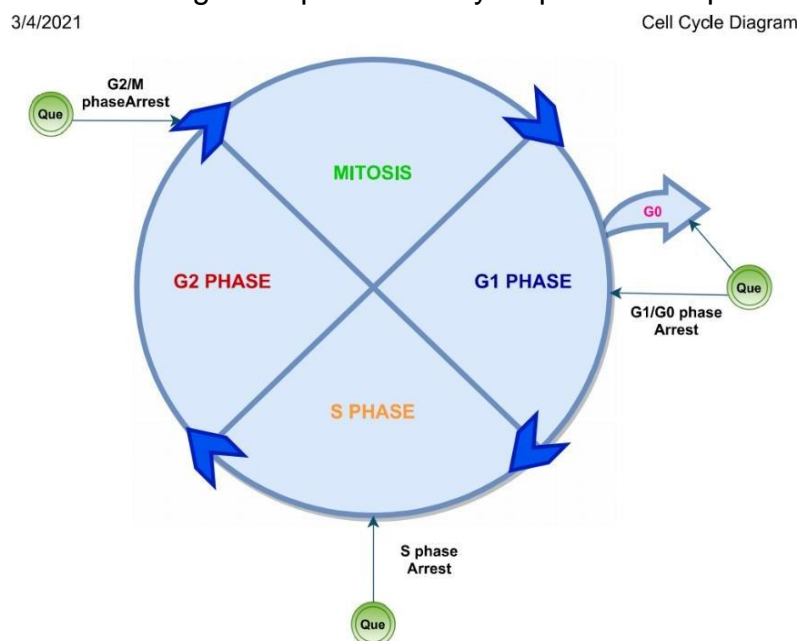
Incubation Time	Risk Ratio (RR)	P value	I <sup>2</sup> index (heterogeneity)
6 hours	0.79	$P<0.0001$	(only one study)
24 hours	0.69 [0.56,0.85]	$P<0.0001$	96%
48 hours	0.67 [0.56, 0.79]	$P<0.0001$	95%

#### Sub-group Analysis on Cell type:

The second Sub-group analysis was performed on the different cell lines (additional data are given in Online resource 1). Eleven studies were evaluated in this analysis. The breast cancer cell lines consisted of; MCF7 [13][21][49][64][87], MDAMB231[93][47][78][87], MDAMB157[78] and SKBR3[94]. Results of the meta- analysis revealed that MCF7 cells with an RR value of 0.65 [0.53,0.81] had the greatest percentage decrease compared to the other cell lines. The MDAMB157 cell line resulted in an RR value of 0.90 [0.84,0.96] showing the least effects of quercetin on percentage cell viability (additional data are given in Online resource 1).

**Table 3:** Summarized Subgroup Meta-Analysis Results – Cell Type = Subgroup

Cell Type	Risk Ratio	P value	I <sup>2</sup> index (Heterogeneity)
MCF 7	0.65 [0.53,0.81]	$P<00001$	92%
MDAMB231	0.78 [ 0.63, 0.96]	$P<00001$	94%
MDAMB157	0.90 [0.84,0.96]	$P<0.02$	Only one study
SKBR3	0.74 [0.66,0.83]	$P<0.03$	Only one study

**Figure 3** Illustration demonstrating the reported cell cycle points that quercetin induces arrest in

**Fig 3:** Illustration demonstrating the different cell cycle checkpoints quercetin induces arrest at. One report shows cell cycle arrest at S phase <sup>[13]</sup>; whereas another described cell cycle arrest at G2/M phase <sup>[93]</sup>. There have also been reports that quercetin induced cell cycle arrest at both phases <sup>[58]</sup>. Xu et al., 2020 showed G1/G0 arrest.

## Discussion

Quercetin has been reported to possess cytotoxic abilities in various cancers <sup>[22][44][45][46][59]</sup>. Previous literature reviews have been published regarding the anti-cancer effects of quercetin in breast cancer <sup>[25][71]</sup> however majority have been qualitative and lack the rigorous systematic approach that allows for evidence-based conclusions <sup>[17]</sup>. More recently there has been a systematic review of the effect of quercetin on MCF-7 and MDA-MB-231 cells. This systematic review and meta-analysis evaluate the efficacy of the cytotoxic effects induced by quercetin in more breast cancer cells lines with the aim to further knowledge surrounding its functional features as an anti-cancer agent and to progress to more *in vivo* studies and clinical trials. Fourteen articles were reviewed. Crucially, all studies reported a reduction in cell growth, proliferation, and apoptosis induction, however these effects are dependent on concentration, incubation time and cell type studied.

Cell viability was chosen as an indicator of cytotoxicity as the consistent method observed across the majority of papers was the 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. This was chosen in consideration of the quantitative aspect of the meta-analysis in order to reduce variations across studies. Among the selected articles, cell viability was evaluated using; MTT assay by ten studies <sup>[93][16][20][33][43][47][49][58][64][78]</sup>, a Cell counting Kit-8 (CCK-8) assay <sup>[87]</sup> and the Trypan blue exclusion assay <sup>[13][21][104]</sup> (table 1). It should be noted that the trypan blue assay is the only one that measures the number of viable cells whereas the other two methods measure mitochondrial metabolism <sup>[76]</sup>.

Seven studies reported a dose and time dependency <sup>[93][13][16][20][33][47][49][58]</sup> on the effects of quercetin whereas three reported a dose dependent <sup>[78][64][43]</sup> response. Seven studies tested concentrations below 50  $\mu\text{M}$ ; 0-25 $\mu\text{M}$  <sup>[13][21][33][47][49][58][87]</sup> and six observed a reduction in cell viability <sup>[13][21][33][47][49][58]</sup> where two

reported statistical significance [13][21]. Interestingly, one study [87] reported a statistically significant increase in percentage cell viability at the concentrations of 5-10  $\mu\text{M}$ . Other studies have also demonstrated biphasic behaviour of quercetin depending on dosage in MCF7 cells [85]. Quercetin concentrations as low as 10 $\mu\text{M}$  have been demonstrated to possess pharmacokinetic relevance in humans [30] thus further *in vitro* research investigating the anticancer effects of quercetin should consider understanding its mechanisms at lower concentrations.

Interestingly, quercetin has shown variations in inducing cell cycle arrest (fig 3). Hence, there is scope to say that the mechanism of action of quercetin may vary according to certain doses and time of incubation. Xu *et al.*, 2020 demonstrated that at high concentrations quercetin promoted G0/G1 arrest whereas lower concentrations induced transition of G1 to S phase. Seven studies reported apoptosis induction through the intrinsic mitochondrial pathway [93][13][21][43][47][64][78] (fig 2). However other apoptotic pathways have also been proposed [87][58][49][944] (fig 2).

Thus, a meta-analysis was conducted to determine the most effective dosage of quercetin for reducing cancer cell viability. Results revealed that increasing doses of quercetin did result in greater decrease cell viability therefore supporting the notion of dose dependency reported by the studies (Table A4). However, contrary to expectations, the differences in RR values of 50 $\mu\text{M}$  and 100 $\mu\text{M}$  were not substantial; 0.79 and 0.74 respectively. In contrast the concentrations greater than 100 $\mu\text{M}$  (120-200  $\mu\text{M}$ ) demonstrated a greater difference in RR value being 0.56. This suggests that the dose range of 50-100  $\mu\text{M}$  may not induce clinically significant results [5]. Nonetheless, this conclusion is limited by the fact that data had to be rounded when inputted into RevMan and this may have affected the statistical interpretation [83]. Furthermore, data obtained from Chou *et al.*, 2010 was also

drastically anomalous, being low value of only 9% cell viability [13] at 24 hours incubation which requires further looking into.

Based on the literature it was evident that time was also a factor that affected the actions of quercetin [93][13][16][20][33][47][49][58]. Hence, a subgroup meta-analysis was performed on the different incubation times presented in the studies. Although the highest incubation time showed the greatest decrease in percentage cell viability; it was relatively similar to the 24-hour subgroup (additional data are given in Online resource 1). This suggests that perhaps incubation beyond 24 hours does not lead to substantial differences and quercetin may have reached saturation in its effects. However concrete conclusions cannot be drawn based on these results alone due to the small range of incubation times included. For instance, Chou *et al.*, 2010 reported apoptosis induction during the 48- hour time point. Nguyen *et al.*, 2017 also noted steady reduction in percentage viability between the times of 24 and 72 hours (additional data are given in Online resource 1).

Given the considerable  $I^2$  index scores and previous reports [95][78][87], it was also speculated that cell type may have been the source of the heterogeneity. A subgroup analysis was performed on all the cell types presented across the studies. The most common cell types used were the MCF7 and the MDAMB231 cell lines which were seen in nine [13][16][21][33][43][47][49][64][87] and six [93][20][47][58][78][87] of the studies, respectively. These 2 cell lines are the most used cell types for laboratory breast cancer research as the MCF7 cell line is typically used as a hormone dependent model whereas the MDAMB231 is typically representative of triple negative [14][37][81]. Furthermore, the majority of studies were Eastern; particularly China [93][16][87], India [20][21][64] and Iran [33][43] that mostly used MCF7 cells whereas the only western study [94] used the SKBR3 cell line therefore it maybe speculated that perhaps certain cell types were more accessible in a particular location. Subgroup meta-analysis results revealed that

quercetin did induce its greatest effects in the MCF7 cell line (additional data are given in Online resource 1). In accordance with this other reported that quercetin acts as an oestrogen receptor antagonist [8][95][51][55]. These findings suggest that perhaps the role of quercetin in breast cancer would be best as an endocrine therapy.

Given the *in Vitro* nature of the studies, a risk of bias assessment using the Cochrane risk of bias reporting could not be used as this is targeted towards clinical studies [38]. Hence the *in vitro* section of the ToxRTool was used to assess reliability of studies used. From the risk of bias assessment (table 2), it can be seen that all of the papers were reliable, however those who had aspects that were classified as unreliable were because a negative control used was not named [20][94][58][49]. Majority of papers used a 0.1% Dimethylsulfoxide (DMSO) solution as a control [13][16][21][4][78][87] whereas others used untreated control cells [43][33]. Comparison of the effects of quercetin on cell viability was done with caution, given the wide variation in concentration ranges across studies. Dilution series of quercetin were conducted from different stock concentrations therefore the same doses may have still differed slightly (either being more diluted or more concentrated) [4].

Implications of this research are that there is enough information surrounding the cytotoxic effects of quercetin to be progressed forward. Although limited, previous *in vivo* studies have confirmed that quercetin does induce cytotoxicity in mice xenografted with breast cancer tumours [33][15]. However, progression to human clinical trials is scarce. Findings of this research postulate that concentrations of quercetin greater than 100µM will lead to clinical significance, therefore this concentration should be transitioned and further investigated in a more dynamic and physiological environment to determine the effects in humans and safety. A potential avenue to explore is the usage of 3D cellular models which will better mimic the

tumour anatomy in a human body and therefore a better insight into the mechanisms of quercetin at different concentrations will be obtained [12][69]. However, animals should be used to observe the safety of quercetin using various doses as carcinogenic effects upon quercetin administration in animal studies have been controversial [32].

### **Conclusion:**

This systematic review and meta-analysis postulate that quercetin has promising potential as an agent for breast cancer treatment. Although further research is warranted in elucidating an optimal dosage that is physiologically relevant; the current literature on cellular studies suggests that quercetin induces cytotoxic effects on human breast cancer cells at high doses and these are determined by incubation time and specific cell type signaling pathways.

### **Declarations**

#### **Ethical approval and consent to participate**

The faculty research ethics committee waived the need for ethics approval and the need to obtain consent for the collection, analysis and publication of this systematic review and meta-analysis

#### **Consent for publication**

Not applicable

#### **Availability of data and materials**

All data generated or analysed during this study are available from the corresponding author on request.

#### **Competing interests**

The authors declare that they have no competing interests

#### **Funding**

The authors received no financial support for the research, authorship, and/or publication of this article

#### **Authors' contribution**

RB contributed to the literature search, data extraction, analysis of the results and writing of

the manuscript. BI designed and directed the systematic review, confirmed accuracy of the literature searches and data interpretation, and revised the manuscript critically for intellectual content. All authors approved the version of the manuscript to be published.

## Acknowledgements

Not applicable

## References:

- [1]. Abdal Dayem, A., Choi, H.Y., Yang, G., Kim, K., Saha, S.K. and Cho, S. (2016) 'The anti-cancer effect of polyphenols against breast cancer and cancer stem cells: molecular mechanisms', *Nutrients*, 8(9), pp. 581. [1]
- [2]. Ahmad, A. (2019) *Breast Cancer Metastasis and Drug Resistance: Challenges and Progress*. Springer. [2]
- [3]. Ahn, J., Lee, H., Kim, S., Park, J. and Ha, T. (2008) 'The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways', *Biochemical and biophysical research communications*, 373(4), pp. 545-549. [3]
- [4]. Ahrar, S., Hwang, M., Duncan, P.N. and Hui, E.E. (2014) 'Microfluidic serial dilution ladder', *Analyst*, 139(1), pp. 187- 190. [4]
- [5]. Andrade, C. (2015) 'Understanding relative risk, odds ratio, and related terms: as simple as it can get', *The Journal of clinical psychiatry*, 76(7), pp. 857-861. [5]
- [6]. Batra, P. and Sharma, A.K. (2013) 'Anti-cancer potential of flavonoids: recent trends and future perspectives', *3 Biotech*, 3(6), pp. 439-459. [6]
- [7]. Brisdelli, F., Bennato, F., Bozzi, A., Cinque, B., Mancini, F. and Iorio, R. (2014) 'ELF-MF attenuates quercetin-induced apoptosis in K562 cells through modulating the expression of Bcl-2 family proteins', *Molecular and cellular biochemistry*, 397(1), pp. 33-43. [7]
- [8]. Brunetti, A. and Manfioletti, G. (2019) 'Hormone Receptors and Breast Cancer', *Frontiers in endocrinology*, 10, pp. 205. [8]
- [9]. Budihardjo, I., Oliver, H., Lutter, M., Luo, X. and Wang, X. (1999) 'Biochemical pathways of caspase activation during apoptosis', *Annual Review of Cell and Developmental Biology*, 15(1), pp. 269-290. [9]
- [10]. Cai, X., Fang, Z., Dou, J., Yu, A. and Zhai, G. (2013a) 'Bioavailability of quercetin: problems and promises', *Current medicinal chemistry*, 20(20), pp. 2572- 2582. [10]
- [11]. Chaicharoenaudomrung, N., Kunhorm, P. and Noisa, P. (2019) 'Three-dimensional cell culture systems as an in vitro platform for cancer and stem cell modeling', *World journal of stem cells*, 11(12), pp. 1065. [12]
- [12]. Chien, S., Wu, Y., Chung, J., Yang, J., Lu, H., Tsou, M., Wood, W.G., Kuo, S. and Chen, D. (2009) 'Quercetin-induced apoptosis acts through mitochondrial-and caspase-3-dependent pathways in human breast cancer MDA-MB-231 cells', *Human & experimental toxicology*, 28(8), pp. 493-503[93]
- [13]. Chou, C., Yang, J., Lu, H., Ip, S., Lo, C., Wu, C., Lin, J., Tang, N., Chung, J. and Chou, M. (2010) 'Quercetin- mediated cell cycle arrest and apoptosis involving activation of a caspase cascade through the mitochondrial pathway in human breast cancer MCF-7 cells', *Archives of Pharmacal Research*, 33(8), pp. 1181-1191. [13]
- [14]. Dai, X., Cheng, H., Bai, Z. and Li, J. (2017) 'Breast cancer cell line classification and its relevance with breast tumor subtyping', *Journal of Cancer*, 8(16), pp. 3131. [14]
- [15]. Dechsupa, S., Kothan, S., Vergote, J., Leger, G., Martineau, A., Beranger, S., Kosanlavit, R., Moretti, J. and Mankhetkorn, S. (2007) 'Quercetin, Siamois 1 and Siamois 2 induce apoptosis in human breast cancer MDA-mB-435 cells xenograft in vivo', *Cancer biology & therapy*, 6(1), pp. 56- 61. [15]
- [16]. Deng, X., Song, H., Zhou, Y., Yuan, G. and Zheng, F. (2013) 'Effects of quercetin on the proliferation of breast cancer cells and expression of survivin in vitro', *Experimental and therapeutic medicine*, 6(5), pp. 1155-1158. [16]
- [17]. Denyer, D. and Tranfield, D. (2009) 'Producing a systematic review.', . [17]
- [18]. Desai, A.G., Qazi, G.N., Ganju, R.K., E El-Tamer, M., Singh, J., Saxena, A.K., Bedi, Y.S., Taneja, S.C. and Bhat, H.K. (2008a) 'Medicinal plants and cancer chemoprevention', *Current Drug Metabolism*, 9(7), pp. 581- 591. [18]
- [20]. Devipriya, S., Vani, G., Ramamurthy, N. and Shyamaladevi, C.S. (2006) 'Regulation of intracellular calcium levels and urokinase activity in MDA MB 231 cells by quercetin', *Chemotherapy*, 52(2), pp. 60-65. [20]
- [21]. Dhumale, S.S., Waghela, B.N. and Pathak, C. (2015) 'Quercetin protects necrotic insult and promotes apoptosis by attenuating the expression of RAGE and its ligand HMGB1 in human breast adenocarcinoma cells', *IUBMB life*, 67(5), pp. 361- 373. [21]
- [22]. Duo, J., Ying, G., Wang, G. and Zhang, L. (2012) 'Quercetin inhibits human breast cancer cell proliferation and induces apoptosis via Bcl-2 and Bax regulation', *Molecular medicine reports*, 5(6), pp. 1453-1456. [22]

- [23]. Elmore, S. (2007) 'Apoptosis: a review of programmed cell death', *Toxicologic pathology*, 35(4), pp. 495-516. [23]
- [24]. Eriksen, M.B. and Frandsen, T.F. (2018) 'The impact of patient, intervention, comparison, outcome (PICO) as a search strategy tool on literature search quality: a systematic review', *Journal of the Medical Library Association: JMLA*, 106(4), pp. 420. [24]
- [25]. Ezzati, M., Yousefi, B., Velaei, K. and Safa, A. (2020) 'A review on anti-cancer properties of Quercetin in breast cancer', *Life Sciences*, 248, pp. 117463. [25]
- [26]. Ferry, D.R., Smith, A., Malkhandi, J., Fyfe, D.W., deTakats, P.G., Anderson, D., Baker, J. and Kerr, D.J. (1996) 'Phase I clinical trial of the flavonoid quercetin: pharmacokinetics and evidence for in vivo tyrosine kinase inhibition.', *Clinical cancer research*, 2(4), pp. 659-668. [26]
- [27]. Fu, Z. and Tindall, D.J. (2008) 'FOXOs, cancer and regulation of apoptosis', *Oncogene*, 27(16), pp. 2312-2319. [27]
- [28]. Gomes, I.B., Porto, M.L., Santos, M.C.L., Campagnaro, B.P., Pereira, T.M., Meyrelles, S.S. and Vasquez, E.C. (2014) 'Renoprotective, anti-oxidative and anti-apoptotic effects of oral low-dose quercetin in the C57BL/6J model of diabetic nephropathy', *Lipids in health and disease*, 13(1), pp. 1-10. [28]
- [29]. Greenwell, M. and Rahman, P. (2015) 'Medicinal plants: their use in anticancer treatment', *International journal of pharmaceutical sciences and research*, 6(10), pp. 4103. [29]
- [30]. Gugler, R., Leschik, M. and Dengler, H.J. (1975) 'Disposition of quercetin in man after single oral and intravenous doses', *European journal of clinical pharmacology*, 9(2), pp. 229-234. [30]
- [31]. Hanahan, D. and Weinberg, R.A. (2000) 'The hallmarks of cancer', *Cell*, 100(1), pp. 57-70. [31]
- [32]. Harwood, M., Danielewska-Nikiel, B., Borzelleca, J.F., Flamm, G.W., Williams, G.M. and Lines, T.C. (2007) 'A critical review of the data related to the safety of quercetin and lack of evidence of in vivo toxicity, including lack of genotoxic/carcinogenic properties', *Food and chemical toxicology*, 45(11), pp. 2179-2205. [32]
- [33]. Hashemzaei, M., Delarami Far, A., Yari, A., Heravi, R.E., Tabrizian, K., Taghdisi, S.M., Sadegh, S.E., Tsarouhas, K., Kouretas, D. and Tzanakakis, G. (2017a) 'Anticancer and apoptosis-inducing effects of quercetin in vitro and in vivo', *Oncology reports*, 38(2), pp. 819-828. [33]
- [36]. Hassan, M., Watari, H., AbuAlmaaty, A., Ohba, Y. and Sakuragi, N. (2014) 'Apoptosis and molecular targeting therapy in cancer', *BioMed research international*, 2014. [36]
- [37]. Holliday, D.L. and Speirs, V. (2011) 'Choosing the right cell line for breast cancer research', *Breast cancer research*, 13(4), pp. 1-7. [37]
- [38]. Hopp, L. (2015) 'Risk of bias reporting in Cochrane systematic reviews', *International journal of nursing practice*, 21(5), pp. 683-686. [38]
- [39]. Huang, C., Chan, C., Chou, I., Lien, C., Hung, H. and Lee, M. (2013) 'Quercetin induces growth arrest through activation of FOXO1 transcription factor in EGFR- overexpressing oral cancer cells', *The Journal of nutritional biochemistry*, 24(9), pp. 1596-1603. [39]
- [40]. Jeong, J., An, J.Y., Kwon, Y.T., Li, L. and Lee, Y.J. (2008) 'Quercetin-induced ubiquitination and down-regulation of Her-2/neu', *Journal of cellular biochemistry*, 105(2), pp. 585- 595. [94]
- [41]. Jørgensen, L., Paludan-Müller, A.S., Laursen, D.R., Savović, J., Boutron, I., Sterne, J.A., Higgins, J.P. and Hróbjartsson, A. (2016) 'Evaluation of the Cochrane tool for assessing risk of bias in randomized clinical trials: overview of published comments and analysis of user practice in Cochrane and non- Cochrane reviews', *Systematic reviews*, 5(1), pp. 1-13. [40]
- [42]. Kanadaswami, C., Lee, L., Lee, P.H., Hwang, J., Ke, F., Huang, Y. and Lee, M. (2005) 'The antitumor activities of flavonoids', *In vivo*, 19(5), pp. 895-909. [41]
- [43]. Kaul, T.N., Middleton Jr, E. and Ogra, P.L. (1985) 'Antiviral effect of flavonoids on human viruses', *Journal of medical virology*, 15(1), pp. 71-79. [42]
- [44]. Khorsandi, L., Orazizadeh, M., Niazvand, F., Abbaspour, M.R., Mansouri, E. and Khodadadi, A. (2017) 'Quercetin induces apoptosis and necroptosis in MCF-7 breast cancer cells', *Bratislava Medical Journal*, 118(2), pp. 123-128. [43]
- [45]. Kim, G.T., Lee, S.H. and Kim, Y.M. (2013) 'Quercetin regulates sestrin 2-AMPK-mTOR signaling pathway and induces apoptosis via increased intracellular ROS in HCT116 colon cancer cells', *Journal of cancer prevention*, 18(3), pp. 264. [44]
- [46]. Kim, H., Moon, J.Y., Ahn, K.S. and Cho, S.K. (2013) 'Quercetin induces mitochondrial mediated apoptosis and protective autophagy in human glioblastoma U373MG cells', *Oxidative Medicine and Cellular Longevity*, 2013. [45]
- [47]. Kim, S., Yoo, E., Woo, J., Han, S., Lee, J., Jung, S., Kim, H. and Jung, J. (2019) 'Antitumor and apoptotic effects of quercetin on human melanoma cells involving JNK/P38 MAPK signaling activation', *European journal of pharmacology*, 860, pp. 172568. [46]

- [48]. Kiyga, E., Şengelen, A., Adıgüzel, Z. and Uçar, E.Ö (2020) 'Investigation of the role of quercetin as a heat shock protein inhibitor on apoptosis in human breast cancer cells', *Molecular biology reports*, 47(7), pp. 4957-4967. [47]
- [49]. Klimisch, H.J., Andreae, M. and Tillmann, U. (1997) 'A Systematic Approach for Evaluating the Quality of Experimental Toxicological and Ecotoxicological Data Regulatory Toxicology and Pharmacology', . [48]
- [50]. Lee, Y., Park, S.Y., Kim, Y., Lee, W.S. and Park, O.J. (2009a) 'AMP kinase/cyclooxygenase-2 pathway regulates proliferation and apoptosis of cancer cells treated with quercetin', *Experimental & molecular medicine*, 41(3), pp. 201-207. [49]
- [51]. Lewis-Wambi, J.S. and Jordan, V.C. (2009) 'Estrogen regulation of apoptosis: how can one hormone stimulate and inhibit?', *Breast cancer research*, 11(3), pp. 1-12. [51]
- [52]. Lindsten, T., Ross, A.J., King, A., Zong, W., Rathmell, J.C., Shiels, H.A., Ulrich, E., Waymire, K.G., Mahar, P. and Frauwirth, K. (2000) 'The combined functions of proapoptotic Bcl-2 family members bak and bax are essential for normal development of multiple tissues', *Molecular cell*, 6(6), pp. 1389-1399. [52]
- [53]. Ma, Y., Yao, C., Liu, H., Yu, F., Lin, J., Lu, K., Liao, C., Chueh, F. and Chung, J. (2018) 'Quercetin induced apoptosis of human oral cancer SAS cells through mitochondria and endoplasmic reticulum mediated signaling pathways', *Oncology letters*, 15(6), pp. 9663-9672. [53]
- [54]. Mihaylova, M.M. and Shaw, R.J. (2011) 'The AMPK signalling pathway coordinates cell growth, autophagy and metabolism', *Nature cell biology*, 13(9), pp. 1016-1023. [54]
- [55]. Miodini, P., Fioravanti, L., Di Fronzo, G. and Cappelletti, V. (1999) 'The two phyto-oestrogens genistein and quercetin exert different effects on oestrogen receptor function', *British journal of cancer*, 80(8), pp. 1150-1155. [55]
- [56]. Mukherjee, A. and Khuda-Bukhsh, A.R. (2015) 'Quercetin down-regulates IL-6/STAT-3 signals to induce mitochondrial-mediated apoptosis in a nonsmall-cell lung- cancer cell line, A549', *Journal of pharmacopuncture*, 18(1), pp. 19. [56]
- [57]. Murota, K., Shimizu, S., Chujo, H., Moon, J. and Terao, J. (2000) 'Efficiency of absorption and metabolic conversion of quercetin and its glucosides in human intestinal cell line Caco-2', *Archives of Biochemistry and Biophysics*, 384(2), pp. 391-397. [57]
- [58]. Nguyen, L.T., Lee, Y., Sharma, A.R., Park, J., Jagga, S., Sharma, G., Lee, S. and Nam, J. (2017) 'Quercetin induces apoptosis and cell cycle arrest in triple-negative breast cancer cells through modulation of Foxo3a activity', *The Korean journal of physiology & pharmacology: official journal of the Korean Physiological Society and the Korean Society of Pharmacology*, 21(2), pp. 205. [58]
- [59]. Niu, G., Yin, S., Xie, S., Li, Y., Nie, D., Ma, L., Wang, X. and Wu, Y. (2011a) 'Quercetin induces apoptosis by activating caspase-3 and regulating Bcl-2 and cyclooxygenase-2 pathways in human HL-60 cells', *Acta Biochim Biophys Sin*, 43(1), pp. 30-37. [59]
- [60]. Noreen, Y., Serrano, G., Perera, P. and Bohlin, L. (1998) 'Flavan-3-ols isolated from some medicinal plants inhibiting COX-1 and COX-2 catalysed prostaglandin biosynthesis', *Planta Medica*, 64(06), pp. 520-524. [61]
- [61]. Ola, M.S., Nawaz, M. and Ahsan, H. (2011) 'Role of Bcl-2 family proteins and caspases in the regulation of apoptosis', *Molecular and cellular biochemistry*, 351(1), pp. 41-58. [62]
- [62]. Pfeffer, C.M. and Singh, A.T. (2018) 'Apoptosis: a target for anticancer therapy', *International journal of molecular sciences*, 19(2), pp. 448. [63]
- [63]. Pradhan, D., Pradhan, R.K., Tripathy, G. and Pradhan, S. (2015) 'Inhibition of proteasome activity by the dietary flavonoid Quercetin associated with growth inhibition in cultured breast cancer cells and xenografts', *Journal of Young Pharmacists*, 7(3), pp. 225. [64]
- [64]. Primikyri, A., Chatziathanasiadou, M.V., Karali, E., Kostaras, E., Mantzaris, M.D., Hatzimichael, E., Shin, J., Chi, S., Briasoulis, E. and Kolettas, E. (2014) 'Direct binding of Bcl-2 family proteins by quercetin triggers its pro- apoptotic activity', *ACS chemical biology*, 9(12), pp. 2737- 2741. [65]
- [65]. Priyadarsini, R.V., Murugan, R.S., Maitreyi, S., Ramalingam, K., Karunakaran, D. and Nagini, S. (2010) 'The flavonoid quercetin induces cell cycle arrest and mitochondria- mediated apoptosis in human cervical cancer (HeLa) cells through p53 induction and NF-κB inhibition', *European journal of pharmacology*, 649(1-3), pp. 84-91. [66]
- [66]. Raina, H., Soni, G., Jauhari, N., Sharma, N. and Bharadvaja, N. (2014) 'Phytochemical importance of medicinal plants as potential sources of anticancer agents', *Turkish Journal of Botany*, 38(6), pp. 1027-1035. [67]
- [67]. Rauf, A., Imran, M., Khan, I.A., ur-Rehman, M., Gilani, S.A., Mehmood, Z. and Mubarak, M.S. (2018) 'Anticancer potential of quercetin: A comprehensive review', *Phytotherapy Research*, 32(11), pp. 2109-2130. [68]
- [68]. Ranganathan, S., Halagowder, D. and Sivasithambaram, N.D. (2015) 'Quercetin suppresses twist to induce apoptosis in MCF-7 breast cancer cells', *PloS one*, 10(10), pp. e0141370.[95]



- [69]. Ravi, M., Paramesh, V., Kaviya, S.R., Anuradha, E. and Solomon, F.P. (2015) '3D cell culture systems: advantages and applications', *Journal of cellular physiology*, 230(1), pp. 16-26. [69]
- [70]. Ravishankar, D., Rajora, A.K., Greco, F. and Osborn, H.M. (2013) 'Flavonoids as prospective compounds for anti-cancer therapy', *The international journal of biochemistry & cell biology*, 45(12), pp. 2821-2831. [70]
- [71]. Reyes-Farias, M. and Carrasco-Pozo, C. (2019) 'The anti- cancer effect of quercetin: molecular implications in cancer metabolism', *International journal of molecular sciences*, 20(13), pp. 3177. [71]
- [72]. Sadhukhan, P., Kundu, M., Chatterjee, S., Ghosh, N., Manna, P., Das, J. and Sil, P.C. (2019) 'Targeted delivery of quercetin via pH-responsive zinc oxide nanoparticles for breast cancer therapy', *Materials science and engineering: C*, 100, pp. 129- 140. [72]
- [73]. Saeidnia, S., Manayi, A. and Abdollahi, M. (2015) 'From in vitro experiments to in vivo and clinical studies; pros and cons', *Current drug discovery technologies*, 12(4), pp. 218- 224. [73]
- [74]. Schulte-Hermann, R., Grasl-Kraupp, B. and Bursch, W. (2000) 'Dose-response and threshold effects in cytotoxicity and apoptosis', *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 464(1), pp. 13-18. [74]
- [75]. Seo, H., Ku, J.M., Choi, H., Choi, Y.K., Woo, J., Kim, M., Kim, I., Na, C.H., Hur, H. and Jang, B. (2016) 'Quercetin induces caspase-dependent extrinsic apoptosis through inhibition of signal transducer and activator of transcription 3 signaling in HER2-overexpressing BT-474 breast cancer cells', *Oncology reports*, 36(1), pp. 31-42. [75]
- [76]. Stoddart, M.J. (2011) 'Cell viability assays: introduction', *Mammalian cell viability*, , pp. 1-6. [76]
- [77]. Su, Q., Peng, M., Zhang, Y., Xu, W., Darko, K.O., Tao, T., Huang, Y., Tao, X. and Yang, X. (2016) 'Quercetin induces bladder cancer cells apoptosis by activation of AMPK signaling pathway', *American journal of cancer research*, 6(2), pp. 498. [77]
- [78]. Sultan, A.S., Khalil, M.I., Sami, B.M., Alkhuriji, A.F. and Sadek, O. (2017) 'Quercetin induces apoptosis in triple- negative breast cancer cells via inhibiting fatty acid synthase and  $\beta$ -catenin', *Int.J.Clin.Exp.Pathol*, 10(1), pp. 156-172. [78]
- [79]. Tang, S., Deng, X., Zhou, J., Li, Q., Ge, X. and Miao, L. (2020) 'Pharmacological basis and new insights of quercetin action in respect to its anti-cancer effects', *Biomedicine & Pharmacotherapy*, 121, pp. 109604. [79]
- [80]. Teekaraman, D., Elayapillai, S.P., Viswanathan, M.P. and Jagadeesan, A. (2019) 'Quercetin inhibits human metastatic ovarian cancer cell growth and modulates components of the intrinsic apoptotic pathway in PA-1 cell line', *Chemico-biological interactions*, 300, pp. 91-100. [80]
- [81]. Theodossiou, T.A., Ali, M., Grigalavicius, M., Grallert, B., Dillard, P., Schink, K.O., Olsen, C.E., Wälchli, S., Inderberg, E.M. and Kubin, A. (2019) 'Simultaneous defeat of MCF7 and MDA-MB-231 resistances by a hypericin PDT- tamoxifen hybrid therapy', *NPJ breast cancer*, 5(1), pp. 1-10. [81]
- [82]. Tinoco, G., Warsch, S., Glück, S., Avancha, K. and Montero, A.J. (2013) 'Treating breast cancer in the 21st century: emerging biological therapies', *Journal of Cancer*, 4(2), pp. 117. [82]
- [83]. Tricker, A.R. (1990) 'The effect of rounding on the significance level of certain normal test statistics', *Journal of Applied Statistics*, 17(1), pp. 31-38. [83]
- [84]. Tummala, R., Lou, W., Gao, A.C. and Nadiminty, N. (2017) 'Quercetin targets hnRNPA1 to overcome enzalutamide resistance in prostate cancer cells', *Molecular cancer therapeutics*, 16(12), pp. 2770-2779. [84]
- [85]. van der Woude, H., Gliszczynska-Świągło, A., Struijs, K., Smeets, A., Alink, G.M. and Rietjens, I.M. (2003) 'Biphasic modulation of cell proliferation by quercetin at concentrations physiologically relevant in humans', *Cancer letters*, 200(1), pp. 41-47. [85]
- [86]. Watters, A.L., Epstein, J.B. and Agulnik, M. (2011) 'Oral complications of targeted cancer therapies: a narrative literature review', *Oral oncology*, 47(6), pp. 441-448. [86]
- [87]. Xu, Z., Zhao, D., Zheng, X., Huang, B., Xia, X. and Pan, X. (2020) 'Quercetin exerts bidirectional regulation effects on the efficacy of tamoxifen in estrogen receptor-positive breast cancer therapy: An in vitro study', *Environmental toxicology*, 35(11), pp. 1179-1193. [87]
- [88]. Yang, T., Kong, B., Gu, J., Kuang, Y., Cheng, L., Yang, W., Xia, X. and Shu, H. (2014) 'Anti-apoptotic and anti-oxidative roles of quercetin after traumatic brain injury', *Cellular and molecular neurobiology*, 34(6), pp. 797-804. [88]
- [89]. Yoshida, T., Maeda, A., Horinaka, M., Shiraishi, T., Nakata, S., Wakada, M., Yogosawa, S. and Sakai, T. (2005) 'Quercetin induces gadd45 expression through a p53- independent pathway', *Oncology reports*, 14(5), pp. 1299- 1303. [89]
- [90]. Zurrida, S. and Veronesi, U. (2015) 'Milestones in breast cancer treatment', *The breast journal*, 21(1), pp. 3-12. [90]